Dopram®-V (doxapram hydrochloride)

Description

DOPRAM-V (doxapram hydrochloride) is a potent respiratory stimulant. It is unique in its ability to stimulate respiration at dosages considerably below those required to evoke cerebral cortical stimulation.

Advantages

Can be administered intravenously in dogs, cats and horses

May be administered subcutaneously, sublingually (topically) or via umbilical vein in neonatal puppies and either subcutaneously or sublingually (topically) in neonatal kittens

Indications

For Dogs, Cats and Horses:

- 1. To stimulate respirations during and after general anesthesia
- 2. To speed awakening and return of reflexes after anesthesia

For Neonate Dogs and Cats:

- 1. To initiate respirations following dystocia or cesarean section
- 2. To stimulate respirations following dystocia or cesarean section

Dosage & Administration

The action of DOPRAM-V is rapid, usually beginning in a few seconds. The duration and intensity of response depend upon the dose, the condition of the animal at the time the drug is administered, and depth of anesthesia. Repeated doses should not be given until the effects of the first dose have passed and the condition of the patient requires it. Dosage should be adjusted for depth of anesthesia, respiratory volume and rate. Dosage can be repeated in15 to 20 minutes, if necessary.

Features

Stimulates respiration in adult dogs, cats and horses as well as neonate puppies and kittens and accelerates arousal from anesthesia

Doxapram minimizes or prevents the undesirable effects of post-anesthetic respiratory depression or hypoventilation and hastens recovery

Dosage of DOPRAM-V for Intravenous Injection

Dogs and Cats				
Weight of Animal (Ib)	Barbiturate anesthesia: Use 1/8 mL (2.5 mg) to 1/4 mL (5 mg) per lb body weight	Gas anesthesia: Use 1/40 mL (0.5 mg) per lb body weight		
10	1 1/4 mL (25 mg) to 2 1/2 mL (50 mg)	1/4 mL (5 mg)		
20	2 1/2 mL (50 mg) to 5 mL (100 mg)	1/2 mL (10 mg)		
30	3 3/4 mL (75 mg) to 7 1/2 mL (150 mg)	3/4 mL (15 mg)		
50	6 1/4 mL (125 mg) to 12 1/2 mL (250 mg)	1 1/4 mL (25mg)		

Horses		
Weight of Animal (Ib)	Chloral hydrate, chloral hydrate and magnesium sulfate barbiturates:	Inhalation anesthesia halothane, methoxyflurane:
	Use 0.0125 mL (0.25 mg) per lb body weight	Use 0.01 mL (0.20 mg) per lb body weight
100	1 1/4 mL (25 mg)	1 mL (20 mg)
200	2 1/2 mL (50 mg)	2 mL (40 mg)
500	6 1/4 mL (125 mg)	5 mL (100 mg)
1,000	12 1/2 mL (250 mg)	10 mL (200 mg)

Neonate Canine

Doxapram may be administered either subcutaneously or sublingually (topically) or via the umbilical vein in doses of 1 - 5 drops (1 - 5 mg) depending on size of neonate and degree of respiratory crisis.

Technique for umbilical vein administration

When the neonate is presented through the incision of the uterus, placental membrane and fluid are removed from mouth and nose. A clamp is placed across the umbilical cord approximately 1 - 2 inches from abdomen of neonate. The umbilical vein is isolated and the selected dose of doxapram injected directly into the umbilical vein.

Neonate Feline

Doxapram may be administered either subcutaneously or sublingually (topically) in a dose of 1 – 2 drops (1 – 2 mg) depending on severity of respiratory crisis.

Benefits

A rapid acting respiratory stimulant that is safe and effective at low doses

A must for every surgical suite

Caution

For intravenous use only in dogs, cats and horses. May be administered subcutaneously, sublingually (topically) or via umbilical vein in neonatal puppies and either subcutaneously or sublingually (topically) in neonatal kittens. Do not mix with alkaline solutions. DOPRAM-V is neither an antagonist of muscle relaxant drugs nor a specific narcotic antagonist.

Doses of DOPRAM-V should be adjusted to meet the requirements of the situation. Excessive doses may produce hyperventilation, which may lead to respiratory alkalosis. A patent air passageway is essential. Adequate, but not excessive, doses should be used and the blood pressure and reflexes should be checked periodically.

How Supplied

DOPRAM-V is available in 20 mL multiple dose vials of the sterile solution.

NDC 0010-4701-01 – 20 mL multiple dose vial – 20 mg/mL Store at controlled room temperature 15 – 30°C (59 – 86°F). Note: Vial should be discarded after a maximum of 50 entries.

HIGHLIGHTS

- Rapid acting respiratory stimulant that is safe and effective at low doses
- Can increase survival of canine and feline neonates in respiratory crisis
- 20 mL multiple dose vials that can be stored at room temperature

Product Information



Ingelheim

5020H

NADA 34-879, Approved by FDA Dopram[®]-V

(doxapram hydrochloride) Injectable, 20 mg/mL

Caution: Federal law restricts this drug to use by or on the order of a licensed veterinarian

Each 1 mL contains:

Doxapram hydrochloride		
Benzyl alcohol		
(as preservative)	0.9%	
Water for injection, USP	q.s.	

Description: Dopram-V (doxapram hydrochloride) is a potent respiratory stimulant. It is unique in its ability to stimulate respiration at dosages considerably below those required to evoke cerebral cortical stimulation. In nonanesthetized animals the dose required to produce convulsions is some 70 to 75 times the dose required to convulsions is some 70 to 75 times ine dose required to produce respiratory stimulation. In an esthetized subjects, doxapram also exerts a marked arousal effect. Thus, by promoting the restoration of normal ventilation and producing early arousal following general anesthesia, doxapram minimizes or prevents the undesirable effects of post-anesthetic respiratory depression or horocarefliction and hastness recovery. hypoventilation and hastens recovery.

Chemistry: The chemical name of doxapram hydrochloride is 1-ethyl-4-(2-morpholinoethyl)-3, 3-diphenyl-2-pyrrolidinone hydrochloride hydrate

The material is prepared as a clear, colorless, 2% aqueous solution with a pH of 3.5 to 5 and is stable at room temperature. Stability studies of 24 months' duration have shown doxapram to have excellent stability characteristics. The preservative is benzy alcohol 0.9% and sterilization is accomplished by aseptic filtration technique. Doxapram is compatible with 5% and 10% dextrose in water or normal saline, but is physically incompatible with alkaline solutions, such as 2.5% thiopental sodium.

Species Variation²⁴⁵

The dog responds more dramatically to doxapram than other species. For example, arousal was not observed in the rat, and the cat responded poorly in comparison with the dog. Respiratory stimulation was slight in the rat, moderate in the cat and marked in the dog and horse.

Effect on EEG

Studies show that while the drug acted selectively on respiratory centers of the brain, higher doses stimulated other parts of the neuraxis. The cortex appeared to be the most resistant part of the central nervous system to the action of the drug.

Effect on Cerebral Blood Flow⁴ The effect of doxapram on cerebral blood flow in The effect of doxapram on cerebral blood flow in anesthetized dogs was determined. Initially, the drug caused a transient increase in blood flow concomitant with rising femoral arterial blood pressure. Flow then diminished while the blood pressure remained elevated. The decreased flow appeared to coincide with marked respiratory stimulation; its occurrence, therefore, is consistent with the known vasoconstrictor effect of bmocramia. hypocaphia.

Effect on Pituitary-Adrenal Axis¹ Intravenous administration of doxapram (20 mg/kg) to anesthetized dogs resulted in a marked rise in the adrenal venous blood level of 17-hydroxycorticosteroids. The peak response occurred at 5-7 minutes in most animals. Hypophysectomy prevented this effect of doxapram.

Site and Mechanism of Action^{2,3,4,7} Doxapram appeared to stimulate respirations primarily by an effect on the brain stem, since sectioning of reflex pathways did not abolish its action. The detection of increased electrical activity in both the inspiratory and expiratory centers of the medulla, at doses as low as 0.2 mg/kg constituted confirmation of this site of action. Only after higher doses were other parts of the brain and spinal cord stimulated. Also, cross circulation experiments have shown that doxapram acts mainly through direct stimulation of central respiratory centers.

through direct sumulation or central respiratory centers. The pressor response to doxaprim appears to be primarily due to stimulation of brain stem vasomotor areas and it is mediated through the sympathoadrenal system. Adrenalectomy and/or drugs which inhibit transmission at sympathetic ganglia or at sympathetic neuroeffector sites were capable of reducing the pressor response to doxapram. Spinal section at C2 abolished the pressor effect pressor effect.

. Intravenous infusion of doxapram to dogs resulted in a prompt and marked increase in total blood and urinary catecholamines.

Therapeutic Ratio

Doxapram did not produce convulsions as readily as did other respiratory stimulants. In unanesthetized animals the ratios between convulsant and respiratory stimulant does of several such drugs were as follows: doxapram, 70; ethamivan, 35; bemegride, 15; pentylenetetrazol, 4; and picrotoxin, 2.3. In animals anesthetized with barbiturates, it was not possible to establish this ratio for doxapram because convulsions could not be produced.

Interaction with Other Drugs^{2,50,10,13} The respiratory stimulant effects of doxapram in dogs were not blocked by anesthetic doses of the following:

Boehringer Ingelheim Vetmedica, Inc. 2621 North Belt Highway St. Joseph, MO 64506-2002 Information 1-866-638-2226 Sales Service 1-800-325-9167 Fax 1-800-318-8366

phenobarbital sodium, pentobarbital sodium, thiopental

social and a social period of the social social social of the social social and the social social and the social s species in which morphine is known to be convulsant.

The respiratory stimulant effects of doxapram in horses were not blocked by anesthetic doses of the following: chloral hydrate, chloral hydrate plus magnesium sulfate and pentobarbital sodium.

Nialamide potentiated the respiratory stimulant action of docapram in dogs and reserpine suppressed this action. In currized dogs, the respiratory response varied inversely with the degree of muscle relaxation existing at the time docapram was administered.

Doxapram antagonized the depressant effects of chlorpromazine, mephenesin and methocarbamol on spinal reflexes in unanesthetized cats.

Various combinations of analeptics in acute barbiturate narcosis in dogs have been compared, including metaraminol and phenylephrine, methetharimide and amphetamine, methetharimide and phenylephrine, amphetamine, methetharimide and phenylephrine, pentylenetetrazol and phenylephrine, pentylenetetrazol and amphetamine, doxapram and phenylephrine, and doxapram alone. While most combinations improved respiratory minute volume quickly, doxapram gave the best response of all. In a similar study comparing the effects of doxapram and various analeptic combinations in dogs, only doxapram was conspicuously effective in increasing ventilation and in shortening sleeping time.

Absorption, Distribution and Fate

Respiratory stimulation was observed in the anesthetized dog after administration by the following routes: intravenous, intramuscular, intraperitoneal, oral, sublingual and subcutaneous.

Spectrophotometric methods were applied to the Spectrophotometric methods were applied to the determination of blood levels and urinary excretion in dogs given doxapram, 10 mg/kg and 20 mg/kg, intravenously. Blood concentrations of doxapram and/or its metabolites were at peak levels immediately after injection and declined rapidly in the first hour. The concentration then further decreased slowly, and an appreciable amount was still present at the end of 24 hours. One dog was given doxapram labeled with radioactive carbon in the 2-position of the pyrrolidinone ring. Blood levels were slightly higher and urinary excretion was slightly lower by isotope assay than by chemical assay. The feces contained 29% of the chemical assay. The feces contained 29% of the administered radioactivity after 48 hours and an additional 9% in the following 24 hours.

Animal Toxicology^{6,9} Oral toxicity studies were carried out in nine dogs and sixty rats for 30 days. Dogs were given down mixe dag and sixty rats for 30 days. Dogs were given dowapram onally by capsule at doses of 20, 50 and 125 mg/kg/day, and one group received the drug intravenously at 20 mg/kg/day. Rats received the drug by stomach tube at 40, 80 and Rats received the drug by stomach tube at 40, 80 and 160 mg/kg/day, with one group receiving 20 mg/kg intravenously daily. Four dogs died, three while receiving the high dose of 125 mg/kg and one at 50 mg/kg. At each dosage level signs of tremor, lacrimation, excessive salivation, occasional vomiting, diarthea, stiffness of the extremities and respiratory stimulation were observed in all dogs. The hemogram, urinalysis and blood chemistry showed no changes which were considered attributable to the drug. the drug.

Histologically, the central nervous system in both species showed congestion, perivascular hemorrhages and petechial hemorrhages. These changes were interpreted as resembling hypoxic changes. The experiments were repeated in does at 2.5, 5, 10 and 20 mg/kg/day and no such lesions were seen.

The acute LD_{50} of doxapram appears to be in the same dose range for various species of animals including mice, rats, adult dogs, newborn dogs and cats. The intravenous $\rm Le_{50}$ was approximately 75 mg/kg while the oral and subcutaneous $\rm Lb_{50}$ is were three to four times greater and the intraperitoneal $\rm Lb_{50}$ about twice as great.

No significant irritation was produced when a saline solution of doxapram at a pH of 4.3 was administered intramuscularly to rabbits at concentrations of 1, 2 and 4%. On the other hand, aqueous solutions of the same concentrations caused tissue irritation in rabbits when given subcutaneously.

Safety Margin for the Various Species'^o The acute LD₂₀ of doxapram HCI in unanesthetized animals appears to be in the same dose range for various species of animals including mice, rats, adult and spected of animats including mice, rats, adult and neonatal dogs and cats. Intravenously, the Lb₂₀ was determined to be approximately 75 mg/kg. The oral and subcutaneous Lb₂₀ was three to four times the intravenous Lb₂₀ whereas the intravenet Lb₂₀ was about twice as great.^{2,9}

The maximum tolerated dose (MTD) of doxapram HCI in unanesthetized animals appears to be in the same dose range for various species of animals including mice, rats, adult and neonatal dogs and cats. Intravenously, the highest MTD tested was determined to be approximately 40 mg/kg. The oral and subcutaneous MTD was three or four times the intravenous MTD whereas the intraperitoneal MTD was about twice as great. Interperturned INTLO Was about thice as great. The highest dose given intravenously to horses was 66 mg per 100 lbs with chloral hydrate anesthesia, and 60 mg per 100 lbs with gas anesthesia. All animals responded normally and no toxic symptoms were observed.

Clinical Studies^{5,12,14}: The clinical use of doxapram in Clinical studies and the clinical use of doxapram in lightly and deeply an esthetical adminish has confirmed the respiratory stimulant and arousal effects previously demonstrated in the laboratory. In one study with 48 dogs and 18 cats subjected to various surgical procedures using pentobarital sodium as the anesthetic, marked increases in ventilation occurred within one minute following a clique laboratory. following a single intravenous injection of 5 mg doxapram per kg of body weight (2.5 mg/lb). The most dramatic improvement occurred in lightly anesthetized dogs pretreated with either promazine or fentanyl-droperidol and atropine. Doxapram accelerated the return of pedal reflexes in all animals.

Doxapram consistently sustained an increased heart rate beginning one minute after injection. A second injection generally failed to further increase heart rate. EKG disturbances of T-wave polarity and magnitude occurred with the use of doxapram but tended to abate with time. Second injections of doxapram generally did not aggravate the EKG distortions.

Ten animals had pre-existing EKG signs of cardiac damage and tolerated doxapram well.

In another study with 73 dogs subjected to various surgical procedures using methoxy-flurane or halothane as the anesthetic, the arousal time was materially shortened, and respiratory minute volume and rate were increased following a single intravenous injection ranging from 0.08 to 1.95 mg/lb with an average dose of 0.44 mg/lb.

Doxapram was effective in intravenous dosages of I mg/kg or less in increasing ventilation and reducing arousal time, especially following methoxyflurane. Tidal volume and respiratory rates were increased; the response normally occurred in 10–20 seconds following injection. No side effects were observed. There were 35 dogs under halothane and 33 dogs under methoxyflurane anesthesia in this study.

In 20 horses subjected to various surgical procedures using intravenous injections of chloral hydrate, chloral hydrate and magnesium sulfate, or pentobarbital as the anesthetic, marked increases in ventilation occurred within 30 seconds following intravenous injection of doxapram in doses ranging from 0.20 to 0.66 mg/lb with an average of 0.28 mg/lb for chloral hydrate and 0.20 to 0.65 mg/lb for the barbiturate. The arousal time was materially detended and reprint proving the yame. materially shortened, and respiratory minute volume and rate were increased.

In another study involving 34 horses anesthetized with halothane or methoxyflurane, marked increases in ventilation occurred within 30 seconds following intravenous injection of doxapram in doses ranging from 0.08 to 0.50 mg/lb, with an average dose of 0.21 mg/lb. The average recovery time was shortened by one-third or more.

In a series of clinical studies involving 80 neonatal canine patients, suffering respiratory crisis following dystocia or caesarean section, doxapram administered either subcutaneously, sublingually or via umbilical vein in doses from 1–5 drops (1–5 mg) resulted in a marked increase in ventilation and survival of all patients.

In a series of clinical studies involving 16 neonatal feline patients, suffering respiratory crisis following caesarean section or dystocia, doxapram administered either subcutaneously or subligically (topically) in doses of 1 to 2 drops (1–2 mg) resulted in a marked increase in ventilation and survival of all patients.

Indications: For Dogs, Cats and Horses: 1. To stimulate respirations during and after general

 anesthesia.
To speed awakening and return of reflexes after anesthesia.

- For Neonate Dogs and Cats: 1. Initiate respirations following dystocia or caesarean section
- 2. To stimulate respirations following dystocia or caesarean section.

Caution: For intravenous use only in dogs, cats and horses. May be administered subcutaneously, sublingually (topically) or via umbilical vein in neonatal pupples and either subcutaneously or sublingually (topically) in neonatal kittens. Do not mix with alkaline solutions. Dopram-V (doxapram hydrochloride) is neither an antagonist of muscle relaxant drugs nor a specific narcotic antagonist.

Doses of Dopram-V should be adjusted to meet the produce hyperventilation Excessive doses may produce hyperventilation which may lead to respiratory alkalosis. A patent air passageway is essential. Adequate, but not excessive, doses should be used and the blood pressure and reflexes should be checked periodically.

DOSAGE OF DOPRAM-V (DOXAPRAM HYDROCHLORIDE) FOR INTRAVENOUS INJECTION Doors and Cats

Dogs and Cats			
Weight of Animal (Ib)	Barbiturate Anesthesia Use ¹ /8 mL (2.5 mg) to ¹ /4 mL (5 mg) per Ib body weight	Gas Anesthesia Use 1/40 mL (0.5 mg) per Ib body weight	
10	1 ¹ / ₄ mL (25 mg) to 2 ¹ / ₂ mL (50 mg)	¹ / ₄ mL (5 mg)	
20	2 ¹ / ₂ mL (50 mg) to 5 mL (100 mg)	¹ / ₂ mL (10 mg)	
30	3 ³ / ₄ mL (75 mg) to 7 ¹ / ₂ mL (150 mg)	³ / ₄ mL (15 mg)	
50	6 ¹ / ₄ mL (125 mg) to 12 ¹ / ₂ mL (250 mg)	11/4 mL (25 mg)	

Dosage should be adjusted for depth of anesthesia. respiratory volume and rate. Dosage can be repeated in 15 to 20 minutes, if necessary.

	Horses	
Weight of Animal (Ib)	Chloral hydrate, chloral hydrate and magnesium sulfate barbiturates, use 0.0125 mL (0.25 mg) per Ib body weight	Inhalation anesthesia halothane, methoxyflurane use 0.01 mL (0.20 mg) per Ib body weight
100 200 500	1 ¹ / ₄ mL (25 mg) 2 ¹ / ₂ mL (50 mg) 6 ¹ / ₄ mL (125 mg)	1 mL (20 mg) 2 mL (40 mg) 5 mL (100 mg)
1000	$12^{1}/_{2}$ ml (250 mg)	10 ml (200 ma)

DOSAGE OF DOPRAM-V (DOXAPRAM HYDROCHLORIDE) FOR NEONATE USE

Neonate Canine

Doxapram may be administered either subcutaneously, sublingually (topically) or via the umbilical vein in doses of 1–5 drops (1–5 mg) depending on size of neonate and degree of respiratory crisis.

Technique for Umbilical Vein Administration

When the neonate is presented through the incision of the uterus, placental membrane and fluid are removed from mouth and nose. A clamp is placed across the umbilical cord approximately 1–2 inches from abdomen of neonate. The umbilical vein is isolated and the selected dose of doxapram injected directly into the umbilical vein.

Neonate Feline

Doxapram may be administered either subcutaneously or sublingually (topically) in a dose of 1–2 drops (1–2 mg) depending on severity of respiratory crisis.

Administration and Dosage: The action of Dopram-V (doxapram hydrochloride) is rapid, usually beginning in a toospharing down on the duration and intensity of response depends upon the dose, the condition of the animal at the time the drug is administered, and depth of anesthesia. Repeated doses should not be given until the effects of the first dose have passed and the condition of the patient requires it.

Dosage should be adjusted for depth of anesthesia, respiratory volume and rate. Dosage can be repeated in 15 to 20 minutes, if necessary.

Note: Vial should be discarded after a maximum of 50 entries

How Supplied: Dopram-V (doxapram hydrochloride) is available in 20 mL multiple dose vials of the sterile solution.

20 mg/mL

Store at controlled room temperature 15 - 30°C (59 - 86°F).

Bibliography

- 1. Lunsford, C.; Cale, Jr., A. D.; Ward, J. W.; Franko, B. V. and Jenkins, H: 4-(b-substituted ethyl)-3, 3-diphenyl-2-pyrrolidinones. A new series of CNS stimulants. J. Med. Chem. 7:302 (1964).
- Ward, J. W. and Franko, B. V.: A New Centrally Acting Agent (AHR-619) with Marked Respiratory Stimulating, Pressor, and "Awakening" Effects; Fed. Proc. 27:(2):325 (1962).
- 3. Funderburk, W. H.; Oliver, K. L. and Ward, J. W.: Electrophysiologic Analysis of the Site of Action of Doxapram Hydrochloride. J. Pharmacol. Exp. Ther. 151:3 (1966).
- 4. Funderburk, W. H.; Oliver, K. L.; Ward, J. W.: Cerebral Blood Flow Changes Due to Doxapram Hydrochloride (AHR-619); Fed. Proc. 22:(2):482 (Abstract) (1963).
- Reports on File. Pharmacology Department, A. H. Robins Company.
- 6. Alphin, R. S. and Franko, B. V.: Inhibition and Stimulation of Gastric Secretions by Doxapram Hydrochloride (AHR-619); Fed. Proc. 22(2):662 (Abstract) (1963).
- Kato, H. and Buckley, J. P.: Possible Sites of Action of the Respiratory Stimulant Effects of Doxapram Hydrochloride. J. Pharmacol. Exp. Ther. 144:260 (1964)
- 8. Bruce, R. B.; Pitts, J. E.; Pinchbeck, F. and Newman, J.: Excretion, Distribution, and Metabolism of Doxapram Hydrochloride. J. of Med. Chem. 8:157 (1965).
- 9. Woodard, G.; Ward, J. W. and Mann, G. T.: Safety Evaluation of the Respiratory Stimulant Doxapram Hydrochloride by Oral and Parenteral Administration to Laboratory Animals. Tox. and Appl. Pharmacol. 6:364 (1964).
- 10. Klemm, W. R.: Physiologic Responses to Equivalent Doses of Doxapram and Various Analeptic Combinations in Acute Barbiturate Narcosis in Dogs. Tox. and Appl. Pharmacol. 8:505 (1966).
- 11. Klemm, W. R.: Evaluation of Effectiveness of Doxapram and Various Analeptic Combinations in Dogs. J. Am. Vet. Med. Assoc. 148:894 (1966). 12. Jensen, E. C. and Klemm, W. R.: Clinical Evaluation of
- an Analeptic, Doxapram, in Dogs and Cats. J. Am. Vet. Med. Assoc. 150(5):516–525 (1967).
- Polak, A. and Plum, F.: Comparison of New Analeptics in Barbiturate-Poisoned Animals. J. Pharmacol. Exp. Ther 145.27 (1964)
- 14. Short, C. E.: Proc. American Animal Hospital Association, Washington, D. C., 1969

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